

The Importance of Measuring Natural Killer Cell Subpopulations, Including CD57, in Health and Disease

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Abstract

Natural killer (NK) cells and cytotoxic T (CD8⁺) cells are two of the most important types of cells in our body that protect it from foreign invaders. Both are cytotoxic cells, but the natural killer cell is part of the innate immune system, while the cytotoxic T cell is one of the major components of adaptive immunity. The most important function shared by these two different cells is the destruction of pathogen-infected and tumor cells. They both do this by releasing bullet-like granules that penetrate the enemy cells, resulting in the death of the infected or cancerous cells. In this review, we look at both cytotoxic T cells and natural killer cells, but pay particular attention to different subpopulations of natural killer cells, including CD56⁺CD16^{dim}, CD56^{dim}CD16⁺, NKT cells, CD57 NK cells, CD57⁺CD16⁺ NK cells, and CD8⁺CD57⁺ T cells, which are a kind of hybrid between a cytotoxic CD8⁺ T cell and a CD57⁺ NK cell. Among these groups, the CD57 subpopulation has been gaining interest. Previously thought to be a marker of inability to proliferate and immunosenescence, CD57 is now regarded as a marker of higher cytotoxic capacity. CD57 cells are now thought of as highly-differentiated long-lived cells that are more efficient in killing their targets than other NK cell subpopulations. In particular, frequencies of CD57-expressing cells in blood and tissues have been correlated with clinical progress in chronic infections or various cancers and with human aging. Overall, the CD57 NK cell subset may play an important role in immune function and disease pathogenesis. We examine all these cells in relation to health and disease, review the mechanisms by which they kill their target cells, and show why it is important to measure them with regard to patients suffering from particular medical conditions, such as viral infections including COVID-19; long COVID; ME/CFS; cancer; autoimmune disease; toxic chemical exposure; chronic alcoholism; chronic pulmonary disease; chronic severe pain; transplantation; acute physical stress; and early pregnancy loss.

1 Introduction

Natural killer (NK) cells and cytotoxic T (CD8⁺, CTL or TC) cells both originate from bone marrow (1, 2). Our bones produce hematopoietic stem cells which differentiate into common myeloid cells and common lymphoid cells. Common myeloid cells develop into cells of the innate immune system, while common lymphoid progenitors develop into cells of the adaptive immune system. While technically considered an innate immune cell like macrophages, neutrophils and the like, NK cells have been discovered to have adaptive properties as well, so that they are now considered a bridge between innate and adaptive immunity (3, 4).

In order for cytotoxic CD8⁺ T cells to kill their enemy target cells, they first have to be activated or primed by antigen-presenting cells, which give them little pieces of the enemy material so that the CD8⁺ T cells can identify their targets. Cytotoxic CD8⁺ T cells are very efficient in the killing of pathogens, particularly viruses that live in the cell. Cells harboring such viruses express on their surface viral peptides bound to major histocompatibility complex 1 (MHC-1) molecules that are detectable by the cytotoxic T cell (5-8).

Conversely, NK cells kill their target cells without any priming or prior activation by other cells. Instead, NK cells basically have a form of friend-or-foe identification system. For the NK cell, the friend identifier is the MHC-1 molecule on the suspect cell's surface (9, 10). Unlike the CD8⁺ cell, which detects the viral peptide bound to the MHC-1 to identify its target, if the NK cell finds an MHC-1 molecule on the surface of a suspect cell, it will not destroy that cell. If an NK cell does not find an intact MHC-1 on a target cell, it will go into kill mode and destroy that cell. However, there is a third scenario wherein a target cell still has an MHC-1 molecule, but also expresses a ligand, such as a viral antigen, that identifies it as an enemy cell. In this case, the ligand overrides the inhibitory "friend" signal from the MHC-1, and the NK cell goes into assassin mode on the target cell (9,10).

Thus, NK cells and CD8⁺ T cells are complementary partners, cytotoxic sidekicks in kicking the butts of enemy cells (11).

2 Natural Killer (NK) Cells and Subpopulations

Natural killer (NK) cells are a subset of lymphocytes which are part of the innate immune system, and are critical for immune surveillance and defense against infection and cancer (9, 10). NK cells contain bullet-like materials called granules which are filled with different highly potent enzymes, earning them the name large granular lymphocytes. By releasing these granules, they can kill viral-infected cells and cancer cells, defending the body against pathogens and tumors (see Figure 1).

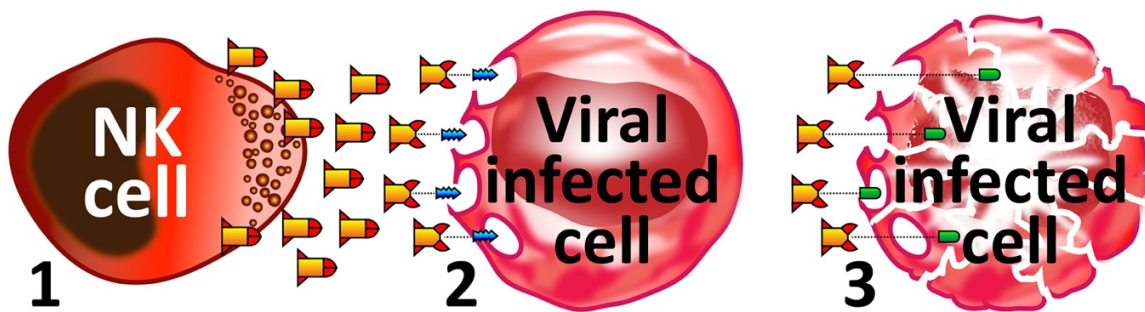


Figure 1. NK bullet attack. 1) The activated NK cell releases the granules (yellow bullets). 2) The granules release perforin (blue bullets), which blast holes into the surface membrane of the viral-infected or cancer cell. 3) The granules then release the granzymes (green bullets), which enter the cell through the holes made by the perforin and detonate inside the cell, causing apoptosis or cell death.

NK cells are involved in immunoregulation, and by producing different cytokines and mediators, they can shape adaptive immune responses (12, 13). Because of this immunoregulatory function, they play a significant role in autoimmunity. NK cells are capable of an array of functions that range widely from their classic anti-tumor and antiviral cytotoxic effector functions to their critical immunoregulatory roles. For example, NK cells activate dendritic cells, macrophages, T cells, T-helper cells, and CD8⁺ cells. NK cells also stimulate plasma cells to produce antibodies (14). These different roles and functions require NK cells to interact with other cells, as shown in Figure 2.

NK cells compose 5 to 15% of peripheral blood mononuclear cells (PBMCs) in humans (9). Humans with deficient or dysfunctional NK cells are susceptible to infections that can potentially be life-threatening (15).

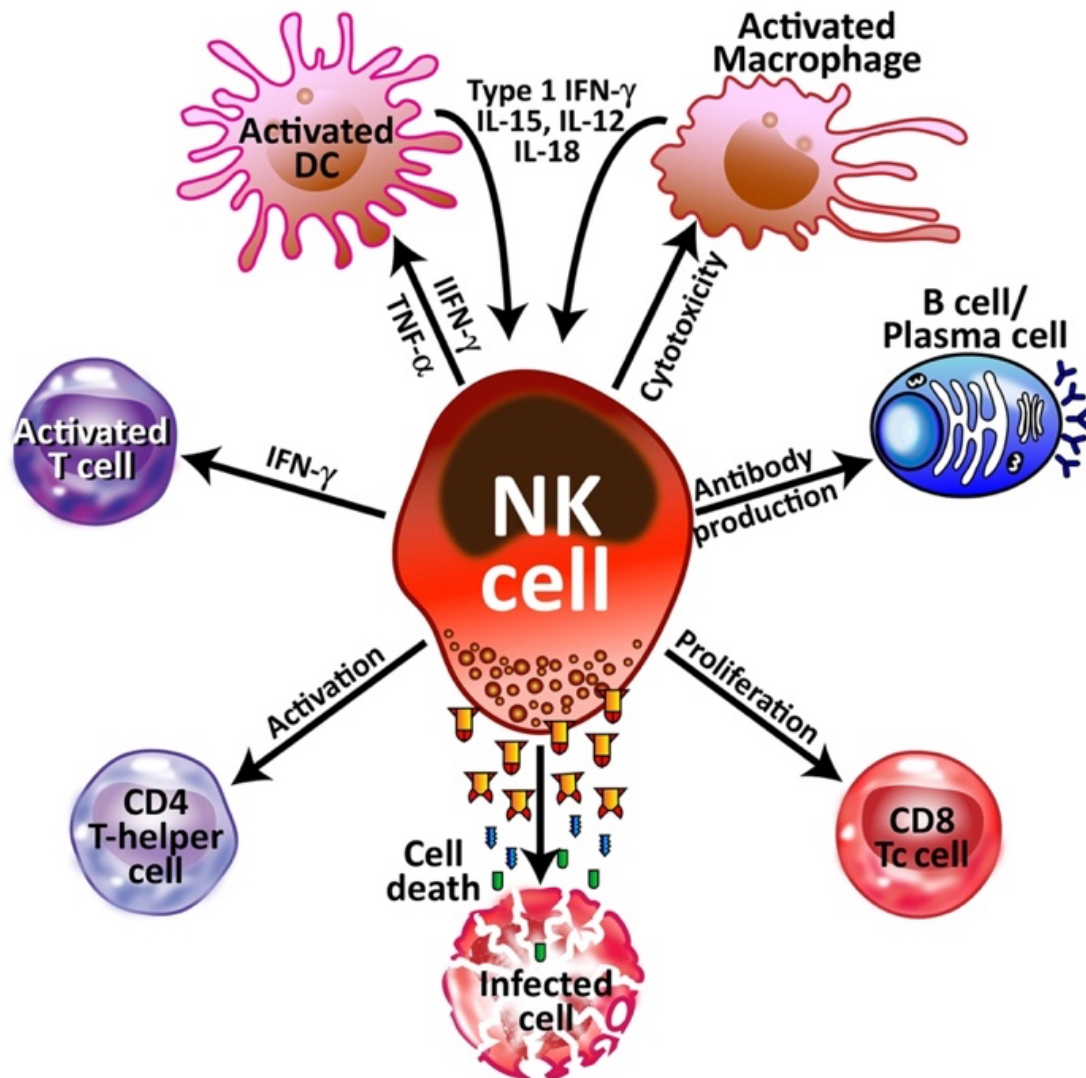
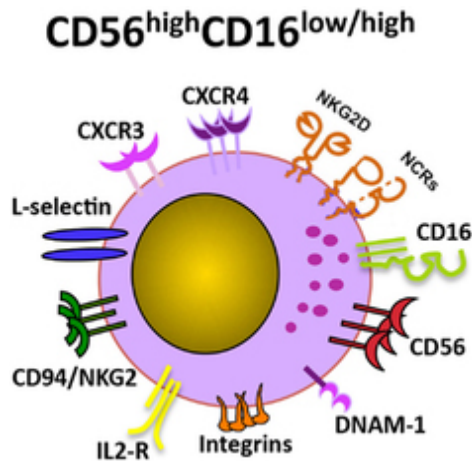


Figure 2. The multiple interactions of NK cells with other cells. NK cells perform various functions and roles that require them to interact with different cells in different ways, from shaping immune responses through the production of cytokines, to the outright killing or destroying of infected cells or cancer cells through the use of bullet-like granules.

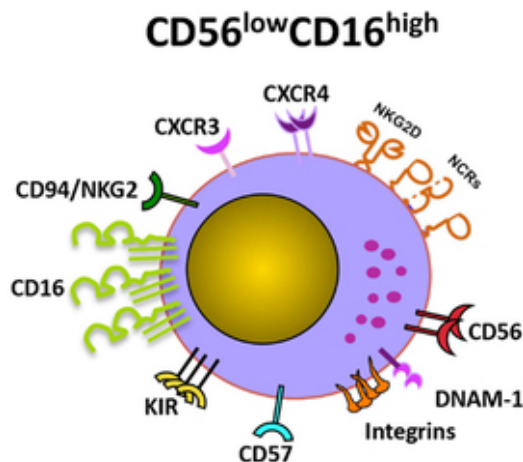
Among the different receptors found on their surfaces, NK cells carry two major cluster differentiation (CD) markers: CD16 and CD56. Based on the amount, strength, and concentration of the markers found on their cell membrane, NK cells can be classified into two basic subsets (16).

2.1 What are CD56⁺CD16^{dim} NK Cells?



CD56^{bright}CD16^{neg/dim} or CD56⁺CD16⁻ NK cells make up about 10% of NK cells and are considered immature. These NK cells are found mainly in the lymphoid tissues. Despite the fact that they are only weakly cytotoxic, they are called regulatory NK cells because they are abundant producers of cytokines and chemokines such as interferon- γ , TNF- β , IL-10, and IL-13, giving them immunoregulatory properties and a role in the shaping of the adaptive immune response. This cytokine-producing ability has been found to be negatively affected in people with myeloma. Phenotypic alterations of CD56⁺ immune cell fraction have been reported in patients with various infectious autoimmune or malignant diseases (16-18).

2.2 What are CD56^{dim}CD16⁺ NK Cells?



CD56^{dim}CD16⁺ or CD56^{low}CD16⁺ NK cells constitute about 90% of circulating NK cells and are considered mature. These are cytotoxic NK cells that are involved in the cytotoxicity of viral-infected cells and tumor cells. They are also involved in cytokine production. CD16 has been described as a receptor expressed on NK cells that facilitates antibody-dependent cellular toxicity (ADCC) by binding to the Fc portion of various antibodies on coated cells. For example, when an antibody binds to a receptor on a tumor cell, the CD56^{low}CD16⁺ NK cell binds to the antibody, allowing the NK cell to release its granules and kill the cell. Likewise, if an anti-EBV antibody binds to a receptor on an EBV-infected cell, the CD56^{low}CD16⁺ NK

cell can bind to that antibody, induce the release of granules, and kill the infected cell (16, 17, 19).

NK cells are further divided into functionally distinct subsets of activating and inhibiting receptors which regulate NK cell activity. Among these are the killer-cell immunoglobulin-like receptors (KIRs), NKG2A, NKG2D, and natural cytotoxicity receptors (20-26).

2.3 What are CD57 Cells?

CD57 is a marker protein found on the surface of a subset of NK cells. They are a highly mature or developed discrete functionally distinct subpopulation of CD16⁺ NK cells that are involved in immunoprotection against pathogens and other environmental factors. Also called memory NK cells, they display high cytotoxic potential. They are highly-differentiated long-lived cells that are thought to be more efficient at killing their targets compared to other NK cell subpopulations. CD57⁺ NK cells have become a matter of recent interest because they are now believed to play a unique role in immune function and have been implicated in a number of health conditions. The CD57 marker is also detected on neural cells, in which it acts as an adhesion molecule, and thus may be involved in the communications between the nervous system and the immune system. CD57⁺ NK cells are

consistently associated with better outcomes in cancer and autoimmune disease. Low levels of CD57⁺ are associated with lower overall survival in cancer (27).

CD57 is a very useful marker of NK cell maturation. Studies show that progression from CD56^{bright} to CD56^{dim}CD57⁻ to CD56^{dim}CD57⁺ reflects a maturation pathway for NK cells (16). Acquisition of CD57 means a higher cytotoxic capacity, greater responsiveness to signaling via CD16 and natural cytotoxic receptors. CD57⁺ cells identify the final stage of NK cell maturation, and their numbers increase with aging (20, 27-29). The numbers of CD57⁺ NK cells also greatly increase in response to chronic exposure to antigens originating from tumor cells, bacteria, and viruses, particularly with EBV, CMV, HIV, hepatitis C and more. Increases in the detectable numbers of CD57⁺ NK cells in blood are also associated with autoimmune disease (20, 30).

In different autoimmune diseases, CD57⁺ NK cells fulfill an immunoregulatory role with their ability to delete autoreactive T cells that are chronically activated by viral antigens. Generally, increases in the population of autoreactive CD57⁺ cells are associated with more severe autoimmune diseases such as Wegener's granulomatosis, multiple sclerosis, type 1 diabetes, Graves' disease, and rheumatoid arthritis. However, there are some instances where certain autoimmune diseases are consistently associated with reduced frequencies or absolute numbers of circulating CD57⁺ NK cells and/or impaired cytotoxicity. In several autoimmune diseases such as atopic dermatitis, Sjögren's syndrome, IgA neuropathy and alopecia areata, a reduction of CD57⁺ NK cells in peripheral blood was reported in comparison to controls (20).

CD57⁺ NK cells are very important in early pregnancy failure. In fact, NK cell counts have been shown to be greater in a subgroup of patients who suffered recurrent pregnancy failure (3.42 ± 2.15) compared to controls (2.14 ± 1.42) (31).

In relation to many pathogens that induce or cause increase in CD57⁺ cells, in 2001 it was reported that there was a reduction in the blood levels of CD57⁺ NK cells in patients with chronic Lyme disease in comparison to those with acute Lyme disease and uninfected individuals (32, 33). However, a 2009 study found no differences between the NK cell counts of patients with chronic Lyme disease and those of controls (34). In fact, since the introduction of CD57 tests in the early 2000s, the first author (AV) has repeatedly stressed that low numbers for CD57 NK cells cannot be used as a specific biomarker for the diagnosis of Lyme disease. In agreement with this position, many authorities such as the CDC (35), North American medical experts (36), European science organizations (37), and the Royal College of Pathologists of Australasia (38) have explicitly warned against the use of CD57 and other unvalidated tests in the diagnosis of Lyme disease. Thus, we measure CD57⁺ cells not for diagnosis of Lyme disease, but for indication of chronic exposure to antigens that originated from bacteria, viruses, tumor cells, and neoantigens formed in the body. The counting of CD57⁺ cells is also recommended for patients with various autoimmune diseases, as a majority of them are induced by environmental factors (20). Finally, we recommend the measurement of CD57⁺ cells for patients who suffered from recurrent pregnancy loss (31).

Overall, CD57⁺ NK cells are a specialized subset of NK cells that may play an important role in immune function and disease pathogenesis (20).

2.3.1 Clinical Importance of Measuring CD57⁺ NK Cells in Peripheral Blood

Persistent antigenic stimulation to various environmental factors, including bacteria, viruses, toxic chemicals and neoantigens results in the alteration of the number of CD57⁺ NK cells.

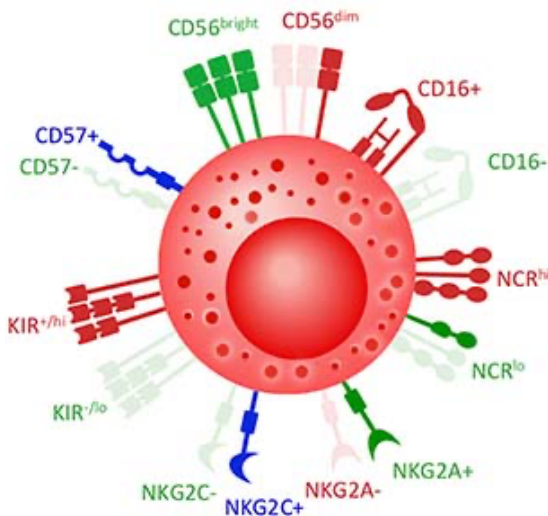
Alterations in CD57⁺ cells have been observed in the following different major disorders (20, 29, 31, 39-44):

- Viral infections including COVID-19
- Long COVID due to viral persistence
- Myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS)
- Cancer
- Autoimmune disease
- Toxic chemical exposure
- Chronic alcoholism due to neoantigen formation
- Chronic pulmonary disease
- Chronic severe pain
- Transplantation
- Acute physical stress
- Early pregnancy loss

This is why the measurement of CD57⁺ NK cells clinically is highly, highly important.

2.3.2 What are CD57⁺CD16⁺ NK Cells?

CD57⁺CD16⁺ Cells



CD57⁺CD16⁺ cells are a subpopulation of CD56^{dim}CD16⁺ NK cells which carry CD57 markers of their surfaces. Cells that carry both CD57 and CD16 markers are highly, highly cytotoxic.

CD57⁺CD16⁺ cells contain the cytolytic enzymes granzyme A, granzyme B and perforin in their granules or bullet-like materials, enabling them to kill cancer cells and virus-infected cells with great efficiency. These cells are considered the most potent cells for combating acute and chronic viral infection (20, 27).

2.3.3 Importance of Measuring CD57⁺CD16⁺ NK Cells in Health and Diseases

1. Increase in CD57⁺CD16⁺ NK cells in chronic viral infection and other pathogens
 - Elevation in CD57⁺CD16⁺ cells is an indication of persistent antigenic stimulation, especially from viruses. Human CMV is one of the clearest examples of infection driving NK cell differentiation, particularly driving the expansion of NKG2C⁺ NK cells, which preferentially acquire CD57. Mature CD57⁺ NK cells expand as a consequence of lifetime exposure to infections, including HBV, HCV, EBV, CMV,

hantavirus, and HIV. Their increase is an indication of ongoing or previous viral infections. Elevation in the levels of CD57⁺ NK cells observed in patients with chronic viral infection correlates with slower disease progression (20, 27).

- Compared to healthy controls, in patients with severe COVID-19, high frequencies of CD57⁺CD16⁺ were detected despite the absolute number of these cells being decreased due to low WBC and low lymphocyte counts. Individuals with long COVID showed significantly increased levels of functional memory cells with high antiviral cytotoxic activity, including NK cells with the CD56⁺CD57⁺NKG2C⁺ phenotype. The hypothesis that a persistent memory cytotoxic response against SARS-CoV-2 could be the cause of long COVID is supported by NK cells expressing both memory (CD57) and activation (NKG2C) markers that did not go away even after the initial SARS-CoV-2 infection was cleared (45-49).

2. Increase in CD57+CD16+ NK cells in cancer

- CD57⁺NK cells are reported in cancer patients. Increased frequency of CD57⁺ NK cells has been correlated with less severe disease and improved survival of patients who suffer from carcinoma, lymphoma, and leukemia. CD57⁺ cells can kill tumor cells through mechanisms that include perforin/granzyme-mediated cytotoxicity (Figure 2) and TRAIL- or Fas-mediated apoptosis (Figure 3). This killing of tumor cells by NK cells is initiated after the binding of the Fas ligand (FasL) or CD95 on the NK cells to the Fas receptor on a tumor cell, thus activating caspases which induce programmed cell death (Figure 3). CD57⁺ and CD16⁺ cells were assessed in patients with oral squamous cell carcinoma (OSCC). High level of CD57⁺ NK cells correlated with longer overall survival. High levels of tumor-associated CD57⁺ NK cells have also been associated with better outcomes in esophageal carcinoma, breast carcinoma, gastric carcinoma, and metastatic carcinoma (50).

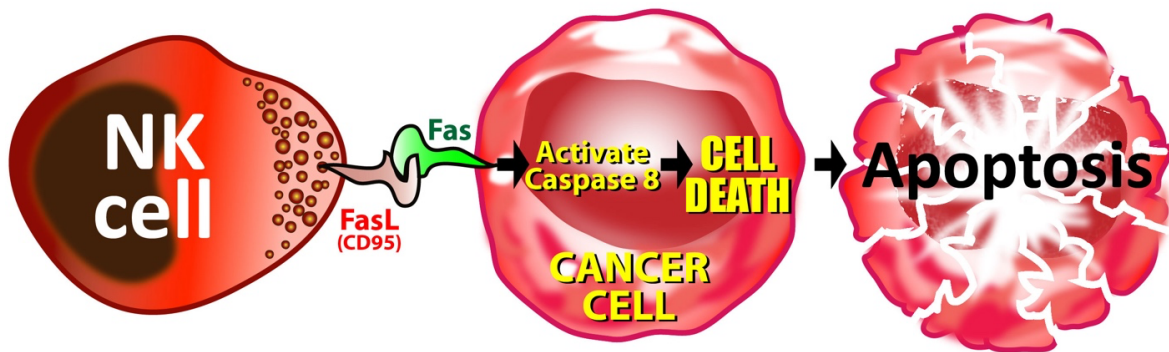


Figure 3. The caspase-mediated pathway to apoptosis. The binding of the CD95 Fas ligand (FasL) on the NK cell to the Fas on the tumor cell initiates a chain of events that activates Caspase 8, ultimately leading to the death or apoptosis of the cancer cell.

3. Changes in CD57+CD16+ NK cells in autoimmune diseases

- Reduced frequencies or absolute numbers of circulating CD57⁺ NK cells and/or impaired NK cell cytotoxicity are consistently associated with autoimmune disease. This supports the regulatory role of cytotoxic CD57⁺ NK cells in preventing or suppressing autoimmune

disease. For example, the blood level of CD57⁺ CD16⁺ cells is depleted in inflammatory and some autoimmune disorders, such as psoriasis, atopic dermatitis, rheumatoid arthritis, and type 1 diabetes. However, an increase in the number of CD57⁺ CD16⁺ cells has been shown in tissues, such as skin, joints and pancreas, of affected individuals. Increased oxidative stress in patients with SLE may have caused preferential apoptosis of mature CD56dimCD57⁺ NK cells, perhaps contributing to the impairment of the ability to eliminate pathogenic CD4⁺ T cells seen in SLE (16, 51).

4. Increase in CD57⁺CD16⁺ NK cells in toxic chemicals

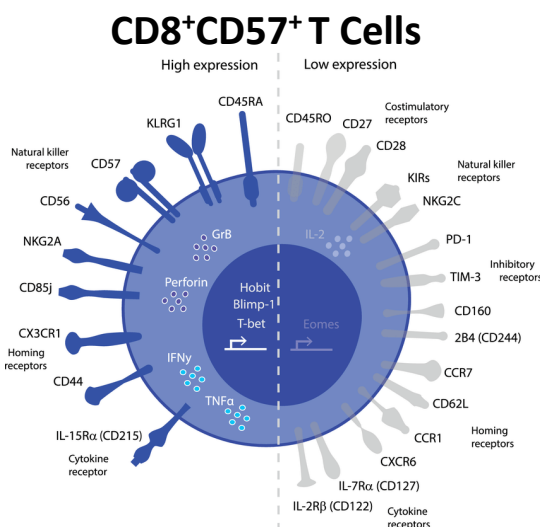
- A significantly higher number of CD57⁺ cells was detected in workers who were chronically exposed to hazardous chemicals such as aromatic amines (benzidine and beta naphthylamine). This increase in CD57⁺ cells in workers exposed to toxic chemicals was associated with a decrease in the number of CD4⁺ lymphocytes (52).

There is another important cell in the body's immune arsenal that partners with the NK cell like a superhero and his sidekick.

3 Cytotoxic CD8⁺ T cells or cytotoxic T lymphocytes (CD8⁺, CTL, TC)

As previously mentioned, CD8⁺ T cells, like NK cells, originate in the bone marrow. However, what makes these cells different is that they migrate to the thymus and develop into maturity there, which is where the "T" in their name comes from. They include cytotoxic T cells, which are the main cellular warriors of the adaptive immune system, specializing in killing virally infected or malignant cells. They are the adaptive counterparts of the innate NK cells (5). They are called cytotoxic lymphocytes because they express cytotoxic molecules such as granzyme A, granzyme B and perforin. They also produce IFN- γ and TNF- α . These functions and properties together enable them to kill viral-infected cells and cancer cells with a high degree of efficiency. They are considered the most potent cells for stopping the spread of pathogens and tumor cells.

3.1 What are CD8⁺CD57⁺ T Cells?



CD8⁺ CD57⁺ T cells are a critical subpopulation of CD8⁺ T cells that are mediators of adaptive immunity. CD8⁺ is formidable enough alone, but putting CD8⁺ together with CD57⁺ to get CD8⁺CD57⁺ is like combining two master killers into one super assassin. There is growing evidence that the CD8⁺CD57⁺ T-cell population plays a significant role in various diseases or conditions, associated with chronic immune activation such as cancer, chronic intracellular infections, chronic alcoholism, some chronic pulmonary diseases, autoimmune diseases, allogeneic transplantation, as well as has a great influence on age-related changes in the immune system status (7).

3.1.1 Importance of Measuring CD8⁺CD57⁺ T Cells in Health and Diseases

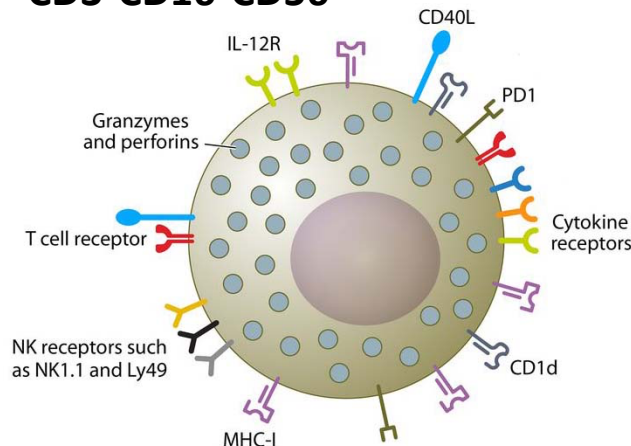
As a result of lifetime exposure to common antigens such as pathogens, neoantigens and autoantigens, the numbers of CD8⁺CD57⁺ cells increase with age. This is because persistent exposure to these antigens induce the expansion of CD8⁺CD57⁺ cells (53, 54).

1. Increase in CD8⁺CD57⁺ cells in chronic viral infection and other pathogens
 - An increase of the CD8⁺CD57⁺ T-cell population is observed in individuals with chronic infections such as EBV, CMV, measles, hepatitis C, parvovirus, HIV, SARS-CoV-2, toxoplasma, and more.
2. Increase in CD8⁺CD57⁺ cells in cancer
 - An increase in the numbers of CD8⁺CD57⁺ T cells has been observed in the blood of patients with different malignancies, including melanomas, head and neck cancer, and hemato-oncological diseases.
3. Changes in CD8⁺CD57⁺ cells in autoimmune diseases
 - Quantitative changes of the CD8⁺CD57⁺ T-cell population are observed in different autoimmune diseases, such as multiple sclerosis, type 1 diabetes, Graves' disease, ankylosing spondylitis, and rheumatoid arthritis. In Graves' disease, ankylosing spondylitis, polymyositis, dermatomyositis and rheumatoid arthritis, increased CD8⁺CD57⁺ T cells are also associated with severity of the disease. In rheumatoid arthritis, after treatment with Abatacept, a decrease of CD8⁺CD57⁺ T cells correlated with clinical response. In other autoimmune diseases, including lupus, type 1 diabetes and multiple sclerosis, decreases in the number of CD8⁺CD57⁺ T cells was observed.
4. Increase in CD8⁺CD57⁺ cells in chronic pulmonary disease
 - The number of CD8⁺CD57⁺ T cells is increased in patients with chronic obstructive pulmonary disease (COPD). This increase is associated with chronic antigenic stimulation coming from pathogens and inhalants which induce injury to epithelial and endothelial cells due to dysregulated immune response.
5. Increase in CD8⁺CD57⁺ cells in transplantation
 - In organ or bone marrow transplantation, the number of CD8⁺CD57⁺ T cells increase significantly to exert immunosuppressive activity. This increase in CD8⁺CD57⁺ T cells in transplant recipients is associated with better graft acceptance and stable function.
6. Increase in CD8⁺CD57⁺ cells in acute physical stress
 - In the case of acute physical stress, including high-intensity physical activity, CD8⁺CD57⁺ T cells are mobilized from peripheral tissue into the blood stream. However, during exercise recovery, these cells will return to their original tissues.
7. Increase in CD8⁺CD57⁺ cells in chronic alcoholism
 - The number of CD8⁺CD57⁺ T cells increases in chronic alcoholism. This increase is most likely caused by hepatocyte damage and chronic antigenic exposure to neoantigens or acetaldehyde-protein adducts after liver parenchymal necrosis. The expansion of CD8⁺CD57⁺ T cells could also be associated with antigens from chronic infections that are characteristic of heavy drinkers.

We have looked at NK cells and CTL cells, but there is a peculiar cell that seems to be half of one and half of the other.

3.2 What are CD3⁺CD16⁺CD56⁺ NKT Cells?

CD3⁺CD16⁺CD56⁺



CD3⁺CD16⁺CD56⁺ cytotoxic natural killer T cells or NKT cells constitute a unique and very rare subset of CD1d-restricted T cells that serve as a bridge between innate and adaptive immunity because they carry receptors characteristic of both T cells and NK cells. They play a significant role at the interface between the innate and adaptive immune systems. They have major modulating effects on immune responses via secretion of cytokines. NKT cells are considered important players in tumor immunosurveillance, and have been shown to play a role in immune protection

against microbial pathogens, different types of cancers, and in the control of autoimmune diseases (55, 56).

4 The Stages of the Cell Death Process

There are many important cell types in the human body, each with a specific function. However, one of the most important cells in our body that protects us against exposome factors is the natural killer (NK) cell. The primary job of the NK cells is the destruction of pathogen-infected cells as well as cancerous cells. They do this by releasing bullet-like materials or cytotoxic granules, which in turn release their own contents to kill the enemy cells.

This cytotoxic action of the NK cell is divided into three main stages:

Stage 1 – Target cell identification

NK cells have been shown to have many antenna-like structures or surface receptors that receive signals indicating there is an enemy nearby. Immediately these surface receptors are activated, not only to identify the signals generated from the surface of the pathogen-infected or cancerous cells, but also to alert the NK cell to prepare its ammunition for the upcoming battle.

Stage 2 – NK cell contact with target cells and formation of immunological synapses

Following target cell identification, the next phase of NK cell action involves NK cell contact with the enemy cells and the formation of cell-to-cell junctions with the target cells. This junction formation between the NK cell and target cells is very important because it ensures that only the harmful enemy cell is destroyed and not an innocent healthy cell. During this synapse formation, the activating signals induce the accumulation and movement of the granules through microtubules formed between the NK cell and the target cell.

Stage 3 – NK-cell-induced target cell death

The last phase of the NK cell's action is the destruction of pathogen-infected cells, cancer cells, cells modified by hazardous chemicals, or other stressed cells. This process which began with the immunological synapse formation is called the degranulation process or release of the bullets. Following the docking process, the granules release perforins that cause pore formation in the surface membrane of the target cell, granzymes, as well as other enzymes, such as serine proteases. Using the pores opened by the perforin, these enzymes gain entry into the enemy cells, where they activate caspase molecules that induce the death of the target cell, also known as apoptosis or the kiss of death. This mechanism by which NK cells kill pathogenic or cancerous cells is shown in Figure 4.

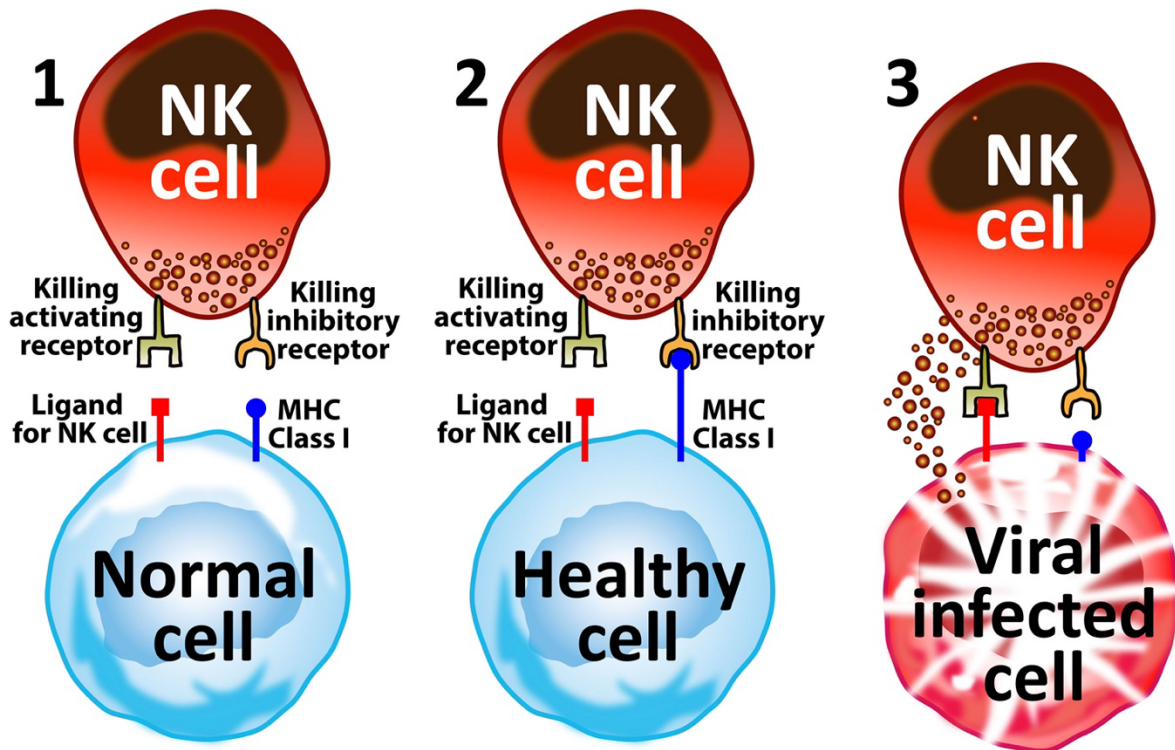


Figure 4. Why NK cells kill viral-infected cells and tumor cells, but not healthy cells. 1. Two different receptors on NK cells and their ligands on healthy cells ensure that NK cells do not attack normal cells. 2. Healthy cells express a significant level of MHC Class I which engages with the killing inhibitory receptor (KIR). Because the ligand for NK cells does not engage the killing activating receptor (KAR), the NK does not kill the healthy cell. 3. In viral-infected and cancer cells, the MHC Class I is significantly reduced, preventing the KIR from engaging with it. But the KAR on the NK cell binds strongly to the ligand for NK cells on the viral-infected/cancer cell, causing the release of destructive missiles from the NK cell, resulting in the killing of the viral-infected cell or tumor cell (see Figures 1, 3).

4.1 How NK cells kill target cells

First, some basics. NK cells are very specific killers. They will only kill infected, harmful or damaged cells. This is possible because NK cells have both a killing inhibitory receptor (KIR) and a killing activating receptor (KAR) (see Figure 4). On the other hand, the body's regular cells have on

the surface membrane a molecule called major histocompatibility complex 1 (MHC-1). These are basically name tags that identify a cell as belonging to you, the host.

If an NK cell comes in contact with a tissue cell and finds a name tag or MHC-1 molecule that identifies it as belonging to the host, the NK cell will leave that cell alone.

But if an NK cell comes in contact with another cell and finds a diminished or no MHC-1 due to viral or tumor damage, then that will be the signal for the NK cell to kill this target cell.

Below are the 5 main ways NK cells use to kill their target cells:

1. Perforin

To perforate means to make a hole, and that's just what perforin does. Once a target cell is identified, the NK cell releases bullet-like materials called cytotoxic granules towards the target. The granules in turn release perforin, which lives up to its name by punching holes into the target cell.

2. Granzymes

The NK cell's cytotoxic granules also release granzymes, which now enter the target through the holes made by the perforin. Once inside the target cell, the granzymes initiate the cell death process by activating the caspases. Think of the caspases as cellular scissors, cutting the DNA, RNA, and vital proteins inside the cell.

3. Fas ligand (FasL)

NK cells also have FS-7-associated surface antigen (Fas) proteins on their surface. If the NK cell's Fas ligand (FasL) comes in contact with a Fas receptor on a target cell, this will initiate the same cell death process that the granzymes do, activating caspases to cut up the insides of target cells.

4. Tumor necrosis factor alpha (TNF- α)

TNF- α is a cytokine, an immune product used by cells to communicate with each other. Similar to FasL, TNF- α can also bind to its own receptor and start the cell death process, destroying the contents of the target cell.

5. Interferon gamma (IFN- γ)

IFN- γ actually does not directly kill harmful cells. However, it can be considered an accessory to cell murder, because what it does is boost immune antibodies and cells like macrophages to turn them into better killing machines. Life Science Identifiers

5 Why Do We Need to Measure CD57⁺ NK Cells, CD57⁺CD16⁺ NK Cells, and CD8⁺CD57⁺ T Cells as Part of the Comprehensive LMAP™ at Cyrex?

- It is recommended for patients who would like to assess their immune system on a yearly basis.

- Individuals with repeated cold and flu and other viral infections.
- Individual who contracted COVID-19, was fully vaccinated, but is still repeatedly contracting the disease.
- Patients exhibiting symptoms of long COVID and/or myalgic encephalomyelitis/chronic fatigue syndrome.
- Person with abnormal blood levels of IgG, IgA, IgM (GAM) and IgG subclasses.
- Person with broken essential barriers (lungs, gut, blood-brain barriers) who are constantly exposed to antigenic stimulation.
- People who are exposed to hazardous chemicals, neoantigens, molds and mycotoxins.
- Individuals in early stages of autoimmunity and full-blown autoimmune disease.
- People suffering from different types of cancer.
- Someone suffering from chronic alcoholism with repeated cold and flu.
- A patient with chronic pulmonary disease.
- Women who have suffered early pregnancy loss.
- Individuals suffering from stress, including acute physical stress.

6 References

1. Bozzano F, Perrone C, Moretta L, De Maria A. NK cell precursors in human bone marrow in health and inflammation. *Front Immunol* (2019) 10:02045. doi: [org/10.3389/fimmu.2019.02045](https://doi.org/10.3389/fimmu.2019.02045)
2. Rothenberg EV, Kueh HY, Yui MA, Zhang JA. Hematopoiesis and T-cell specification as a model developmental system. *Immunol Rev* (2017) 271(1):72-97, doi: [10.1111/imr.12417](https://doi.org/10.1111/imr.12417)
3. Basilio-Queiros D, Mischak-Weissinger E. Natural killer cells – from innate cells to the discovery of adaptability. *Front Immunol* (2023) 14:1172437. doi: [10.3389/fimmu.2023.1172437](https://doi.org/10.3389/fimmu.2023.1172437).
4. Vivier E, Raulet DH, Moretta A, Caligiuri MA, Zitvogel L, Lanier LL, et al. Innate or Adaptive Immunity? The example of natural killer cells. *Science* (2011) 331(6013):44-49. doi: [10.1126/science.1198687](https://doi.org/10.1126/science.1198687).
5. Cassioli C, Baldari CT. The expanding arsenal of cytotoxic T cells. *Front Immunol* (2022) 13:883010. doi: [10.3389/fimmu.2022.883010](https://doi.org/10.3389/fimmu.2022.883010)

6. Raskov H, Orhan A, Christensen JP, Gögenur I. Cytotoxic CD8⁺ T cells in cancer and cancer immunotherapy. *Br J Cancer* (2021) 124(2):359-367. doi: 10.1038/s41416-020-01048-4.
7. Strioga M, Pasukoniene V, Characiejus D. CD⁺ CD28⁻ and CD8⁺ CD57⁺ T cells and their role in health and disease. *Immunol* (2011) 134(1):17-32, 10.1111/j.1365-2567.2011.03470.x
8. Jaime-Sanchez P, Uranga-Murillo I, Aguilo N, Khouili SC, Arias MA, Sancho D, et al. Cell death induced by cytotoxic CD8⁺ T cells is immunogenic and primes caspase-3-dependent spread immunity against endogenous tumor antigens. *J Immunother Cancer* (2020) 8(1):e000528. doi: 10.1136/jitc-2020-000528.
9. Paul S, Lal G. The molecular mechanism of natural killer cells function and its importance in cancer immunotherapy. *Front Immunol* (2017) 8:1124. doi: 10.3389/fimmu.2017.01124.
10. Lanier LL. Up on the tightrope: natural killer cell activation and inhibition. *Nat Immunol* (2008) 9(5): 495–502. doi: 10.1038/ni1581.
11. Rosenberg J, Huang J. CD8⁺ T cells and NK cells: parallel and complementary soldiers of immunotherapy. *Curr Opin Chem Eng* (2018) 19:9-20. doi: 10.1016/j.coche.2017.11.006.
12. Biron CA. Yet another role for natural killer cells: cytotoxicity in immune regulation and viral persistence. *PNAS* (2012) 109(6):1814-1815. doi: 10.1073/pnas.1120528109.
13. Hill TM, Bezbradica JS, Kaer LV, Joyce S. CD1d-restricted natural killer T cells. 2016, in: *Encyclopedia of Life Sciences*. John Wiley & Sons, Ltd: Chichester, doi: 10.1002/9780470015902.a0020180.pub2
14. Poxnanski SM, Ashkar AA. What defines NK cell functional fate: phenotype or metabolism? *Front Immunol* (2019) 10:1414. doi: 10.3389/fimmu.2019.0141
15. Orange JS. Natural killer cell deficiency. *J Allergy Clin Immunol* (2013) 132(3):515–526. doi: 10.1016/j.jaci.2013.07.020.
16. Poli A, Michel T, Thérésine M, Andres E, Hentges F, Zimmer J. CD56^{bright} natural killer (NK) cells: an important NK cell subset. *Immunol* (2009) 126(4):458-465, doi: 10.1111/j.1365-2567.2008.03027.x
17. Gismondi A, Stabile H, Nisti P, Santoni A. Effector functions of natural killer subsets in the control of hematological malignancies. *Front Immunol* (2015) 6:567. doi: 10.3389/fimmu.2015.00567
18. Wang R, Jaw JJ, Stutzman, NC, Zou Z, Sun PD. Natural killer cell-produced IFN- γ and TNF- α induce target cell cytolysis through upregulation of ICAM-1. *J Leukoc Biol* (2012) 91(2):299-309. doi: 10.1189/jlb.0611308.
19. Chijioke O, Landtwing V, Münz C. NK cell influence on the outcome of primary Epstein-Barr virus infection. *Front Immunol* (2016) 7:323, doi: 10.3389/fimmu.2016.00323

20. Nielsen CM, White MJ, Goodier MR et al. Functional significance of CD57 expression on human NK cells and relevance to disease. *Front Immunol*, 2013, 4:422, doi: 10.3389/fimmu.2013.00422
21. Marcenaro M, Notarangelo LD, Orange JS, Vivier E. Editorial: NK cell subsets in health and disease: new developments. *Front Immunol* (2017) 8:1363. doi: 10.3389/fimmu.2017.01363.
22. Walter L, Petersen B. Diversification of both KIR and NKG2 natural killer cell receptor genes in macaques – implications for highly complex MHC-dependent regulation of natural killer cells. *Immunol* (2017) 150(2):139-145. doi: 10.1111/imm.12666.
23. Meckawy GR, Mohamed AM, Zaki WK, Khattab MA, Amin MA, ElDeeb MA, et al. Natural killer NKG2A and NKG2D in patients with colorectal cancer. *J Gastrointest Oncol* (2019) 10(2):218-225. doi: 10.21037/jgo.2018.12.13.
24. Hudspeth K, Silva-Santos B, Mavillo D. Natural cytotoxicity receptors: Broader expression patterns and functions in innate and adaptive immune cells. *Front Immunol* (2013) 4:69. doi: 10.3389/fimmu.2013.00069.
25. Hadad U, Thauland TJ, Martinez OM, Butte MJ, Porgador A, Krams SM. NKp46 clusters at the immune synapse and regulates NK cell polarization. *Front Immunol* (2015) 6:495. doi: 10.3389/fimmu.2015.00495.
26. Björkström NK, Riese P, Heuts F et al. Expression patterns of NKG2A, KIR, and CD57 define a process of CD56^{dim} NK-cell differentiation uncoupled from NK-cell education. *Blood*, 2010, 116(19): 3853-3864, doi: 10.1182/blood-2010-04-281675
27. Lopez-Vergés S, Milush JM, Pandey S et al. CD57 defines a functionally distinct population of mature NK cells in the human CD56^{dim}CD16⁺ NK-cell subset. *Blood*, 2010, 116(19): 3865-3874, doi: 10.1182/blood-2010-04-282301
28. Muntasell A, Servitjia S, Cabo M et al. High numbers of circulating CD57⁺ NK cells associate with resistance to HER2-specific therapeutic antibodies in HER2⁺ primary breast cancer. *Cancer Immunol Res*, 2019, 7(8):1280-1292, doi.org/10.1158/2326-6066.CIR-18-0896
29. Chattopadhyay PK, Betts MR, Price DA et al. The cytolytic enzymes granzyme A, granzyme B, and perforin: expression patterns, cell distribution, and their relationship to cell maturity and bright CD57 expression. *J Leukoc Biol*, 2009, 85(1): 88-97, doi: 10.1189/jlb.0208107
30. Björkström NK, Strunz B, Ljunggren H-G. Natural killer cells in antiviral immunity. *Nat Rev Immunol*, 2022, 22(2): 112-123, doi: 10.1038/s41577-021-00558-3
31. Ozcimen EE, Kiyici H, Uckuyu A et al. Are CD57⁺ natural killer cells really important in early pregnancy failure? *Arch Gynecol Obstet*, 2009, 493-497, doi: 10.1007/s00404-008-0736-y
32. Stricker RB, Winger EE. Decreased CD57 lymphocyte subset in patients with chronic Lyme disease. *Immunol Lett*, 2001, 76(1):43-48, doi: 10.1016/S0165-2478(00)00316-3

33. Stricker RB, Burrascano J, Winger E. Longterm decrease in the CD57 lymphocyte subset in a patient with chronic Lyme disease. *Ann Agric Environ Med*, 2002, 9(1):111-113
34. Marques A, Brown MR, Fleisher TA. Natural killer cell counts are not different between patients with post-Lyme disease syndrome and controls. *Clin Vaccine Immunol*, 2009, 16(8):1249-1250, doi: 10.1128/CVI.00167-09
35. Laboratory tests and practices that are not currently recommended.
<https://www.cdc.gov/lyme/diagnosistesting/labtest/otherlab/index.html>
36. Lantos PM, Rumbaugh J, Bockenstedt LK et al. Clinical practice guidelines by the Infectious Diseases Society of America (IDSA), American Academy of Neurology (AAN), and American College of Rheumatology (ACR): 2020 guidelines for the prevention, diagnosis and treatment of Lyme disease. *Clin Infect Dis*, 2021, 72(1): e1-e48, doi: 10.1093/cid/ciaa1215
37. Mygland A, Ljostad U, Fingerle V et al. EFNS guidelines on the diagnosis and management of European Lyme neuroborreliosis. *Eur J Neurol*, 2010, 17: 8-16, doi:10.1111/j.1468-1331.2009.02862.x
38. The Royal College of Pathologists of Australasia. Position statement: Diagnostic laboratory testing for Lyme disease (or similar syndromes) in Australia and New Zealand. *Microbiology AC*, 2022, 1/2014.
39. Taghavi N, Bagheri S, Akbarzadeh A. Prognostic implication of CD57, CD16 and TGF- β expression in oral squamous cell carcinoma. *J Oral Pathol Med*, 2016, 45(1): 58-62, doi: 10.1111/jop.12320
40. Stabile H, Fionda C, Gismondi A et al. Role of distinct natural killer cell subsets in anticancer response. *Front Immunol*, 2017, 8:293, doi: 10.3389/fimmu.2017.00293
41. Lu Z, Tian Y, Bai Z et al. Increased oxidative stress contributes to impaired peripheral CD56^{dim}CD57⁺ NK cells from patients with systemic lupus erythematosus. *Arthritis Res Ther*, 2022, 24(1):48, doi: 10.1186/s13075-022-02731-y.
42. Song K, Coleman RA, Alber C et al. TH1 cytokine response of CD57⁺ T-cell subsets in healthy controls and patients with alcoholic liver disease. *Alcohol*, 2001, 24(3): 155-167, doi: 10.1016/S0741-8329(01)00146-X
43. Youn J-C, Jung MK, Yu HT et al. Increased frequency of CD4⁺CD57⁺ senescent T cells in patients with newly diagnosed acute heart failure: exploring new pathogenic mechanisms with clinical relevance. *Sci Rep*, 2019, 9: 12887, doi: 10.1038/s41598-019-49332-5
44. Kiu B, Yang G-X, Sun Y et al. Decreased CD57 expression of natural killer cells enhanced cytotoxicity in patients with primary sclerosing cholangitis. *Front Immunol*, 2022, 13: 912961, doi.org/10.3389/fimmu.2022.912961
45. Galán M, Vigón L, Fuentes D et al. Persistent overactive cytotoxic immune response in a Spanish cohort of individuals with long COVID: identification of diagnostic biomarkers. *Front Immunol*, 2022, 13: 848886, doi: 10.3389/fimmu.2022.848886

46. Di Vito C, Calcaterra F, Colaniz N et al. Natural killer cells in SARS-CoV-2 infection: pathophysiology and therapeutic implications. *Front Immunol*, 2022, 13: 888248, doi: 10.3389/fimmu.2022.888248
47. Clavarino G, Leroy C, Epaulard O et al. Fine analysis of lymphocyte subpopulations in SARS-CoV-2 infected patients: differential profiling of patients with severe outcome. *Front Immunol*, 2022, 13: 889613, doi: 10.3389/fimmu.2022.889613
48. Varchetta S, Mele D, Oliviero B et al. Unique immunological profile in patients with COVID-19. *Cell Mol Immunol*, 2021, 18(3): 604-612, doi: 10.1038/s41423-020-00557-9
49. Maucourant C, Filipovic I, Ponzetta A et al. Natural killer cell immunotypes related to COVID-19 disease severity. *Sci Immunol*, 2020, 5(50): eabd6832, doi: 10.1126/sciimmunol.abd6832
50. Gismondi A, Stabile H, Nisti P et al. Effector functions of natural killer subsets in the control of hematological malignancies. *Front Immunol*, 2015, 6: 567, doi: 10.3389/fimmu.2015.00567
51. Michel T, Poli A, Cuapio A et al. Human CD56bright NK cells: an update. *J Immunol*, 2016, 196(7): 2923-2931, doi: 10.4049/jimmunol.1502570
52. Tanigawa T, Araki S, Abo T et al. Increase in CD57 + CD16-lymphocytes in workers exposed to benzidine and beta-naphthylamine: assessment of natural killer cell subpopulations. *Int Arch Occup Environ Health*, 1996, 69(1): 69-72
53. Chijioke O, Landtwing V, Münz C. NK cell influence on the outcome of primary Epstein-Barr virus infection. *Front Immunol*, 2016, 7: 323, doi: 10.3389/fimmu.2016.00323
54. Van den Berg SPH, Pardieck IN, Lanfermeijer J et al. The hallmarks of CMV-specific CD8 T-cell differentiation. *Med Microbiol Immunol*, 2019, 208(3): 365–373, doi: 10.1007/s00430-019-00608-7
55. Hill TM, Bezbradica JS, Van Kaer L et al. CD1d-restricted natural killer T cells. 2016, in: eLS. John Wiley & Sons, Ltd: Chichester, doi: 10.1002/9780470015902.a0020180.pub2
56. Almeida JS, Casanova JM, Santos-Rosa M et al. Natural killer T-like cells: immunobiology and role in disease. *Int J Mol Sci*, 2023, 24(3): 2743, doi: 10.3390/ijms24032743